

Phylogenetic relationships amongst the snake-eyed lizards of the genus *Ablepharus* Fitzinger, 1823 (Sauria, Scincidae) in the Iranian Plateau based on mtDNA sequences

Rasoul Karamiani¹, Nasrullah Rastegar-Pouyani¹, Eskandar Rastegar-Pouyani²

¹ Department of Biology, Faculty of Science, Razi University, Kermanshah, Iran

² Department of Biology, Faculty of Science, Hakim Sabzevari University, Sabzevar, Iran

<http://zoobank.org/0647E59D-F7E1-427F-8EC9-496FC3BFDB85>

Corresponding author: Eskandar Rastegar-Pouyani (rastegarpouyani45@gmail.com)

Academic editor: Peter Mikulíček ♦ Received 23 March 2021 ♦ Accepted 20 June 2021 ♦ Published 15 September 2021

Abstract

We recovered molecular phylogenetic relationships amongst species of the genus *Ablepharus* in Iran and Iraq. Partial sequences of three mitochondrial genes (cytochrome C oxidase subunit I – COI, 12S rRNA and 16S rRNA) were analysed. In addition, phylogenetic relationships and taxonomic evaluation of *Ablepharus* species in Cyprus, India, Greece, Turkey and Syria were performed using partial sequences of the 16S rRNA gene. Phylogenetic trees and estimated genetic distances showed that the *Ablepharus* populations of Iran and Iraq clustered into three distinct clades. One is found in northwest Iran (*A. bivittatus* in Ardabil, East and West Azerbaijan and Hamedan Provinces). The second clade, formed by *A. chernovi*, is found only in Uromia. The third and most heterogeneous clade is divided into two subclades, the first includes two lineages of *Ablepharus* in Khorasan Razavi and Semnan Provinces (*A. pannonicus*) and in eastern and south-eastern Iran (*A. grayanus*); the second subclade is distributed in the eastern part of Iraq and west and south-western Iran (*Ablepharus* sp.). Our analyses indicated that splitting of *A. chernovi* within the genus occurred in the early Miocene [about 22.5 million years ago (Mya)]. *Ablepharus bivittatus* diverged 15.2 Mya, in the middle Miocene. *Ablepharus pannonicus* diverged in the late Miocene (8.4 Mya) and *A. grayanus* separated in the late Miocene (6.7 Mya). The lineages of eastern Iraq and south-western Iran (*Ablepharus* sp.) diverged also in the late Miocene (7.0 Mya).

Key Words

Ablepharus, COI, mitochondrial sequences, 12S rRNA, 16S rRNA, systematics

Introduction

The family Scincidae encompasses more than 25% of all living genera and species of lizards (Uetz and Hošek 2020) with a nearly worldwide distribution (Vitt and Caldwell 2013). Based on morphological characters, they consist of four subfamilies, Acontinae, Feylininae, Lygosominae and Scincinae (Greer 1970), but based on molecular analysis, they include three subfamilies Acontinae, Scincinae and Lygosominae (Pyron et al. 2013). According to the molecular phylogenetic studies, the genus *Ablepharus* is nested within Lygosominae and is sister

taxon of the central and East Asian *Asymblepharus* (Pyron et al. 2013). Despite systematic morphological studies, so far, a limited molecular survey has been done on the genus *Ablepharus* (e.g. Poulakakis et al. 2005; Skourtanioti et al. 2016).

The genus *Ablepharus* Fitzinger, 1823 contains ten valid species: *A. bivittatus* (Ménétries, 1832), *A. budaki* Göçmen, Kumlutaş & Tosunoğlu, 1996, *A. chernovi* Darevsky, 1953, *A. darvazi* Jeremčenko & Panfilov, 1990, *A. deserti* Strauch, 1868, *A. grayanus* (Stoliczka, 1872), *A. kitaibelii* Bibron & Bory, 1833, *A. lindbergi* Wettstein, 1960, *A. pannonicus* (Fitzinger, 1824) and *A. rueppellii*

(Gray, 1839) which are distributed in southern Europe (the whole Balkan Peninsula and islands of the Aegean Sea), Turkey, Syria to Egypt, Azerbaijan, Armenia, Caucasus, Tajikistan, Kazakhstan, Kyrgyzstan, Uzbekistan, Turkmenistan, Afghanistan, Iran, Iraq, United Arab Emirates, Pakistan and NW India (Fühn 1969a, b; Anderson 1999; Khan 2002; Vyas 2011). Skourtanioti et al. (2016), based on two mitochondrial genes (cytochrome *b* and 16S rRNA) and two nuclear markers (the melanocortin 1 receptor – MC1R and the natural killer-tumour recognition – NKTR), found that *A. kitaibelii* represented a species complex that includes species *A. kitaibelii*, *A. budaki*, *A. chernovi* and *A. rueppellii*.

Ablepharus bivittatus, *A. chernovi*, *A. grayanus* and *A. pannonicus* occur in Iran – our study area (Anderson 1999; Karamiani et al. 2015).

The two-streaked snake-eyed skink, *A. bivittatus* was first described as *Scincus bivittatus* from Perimbal, Talysh Mountains and Azerbaijan. *Ablepharus bivittatus* is distributed in eastern Turkey, northern Iran, Armenia and Azerbaijan (Baran and Atatür 1998; Anderson 1999; Ilgaz et al. 2007). It is expected that *A. bivittatus* occurs in higher elevations of the Zagros Mountains and Blanford (1876) mentioned an isolated record from the Lar area in northern Fars Province. A study of phenotypical variation of the populations of Hamedan and East Azerbaijan Provinces from Iran found there was not much difference between them (Karamiani et al. 2017).

Chernov's snake-eyed skink, *A. chernovi*, was regarded as a subspecies of *Ablepharus kitaibelii* from vicinity of the settlement Tkhit, Ashtarak Region, in the middle current of the River Razdan, Armenia. Eremchenko and Szczerbak (1986) wrote a comprehensive monograph about the genus *Ablepharus*. In their study, *A. chernovi*, which had previously been accepted as a subspecies (Fühn 1969a; Baran 1977), is considered a separate species (Schmidtler 1997). A molecular phylogenetic study confirmed *A. chernovi* to represent a genetically-distinct species (Poulakakis et al. 2005). *Ablepharus chernovi* is distributed in Razdan River Valley, Armenia, northern Syria and southern and central parts of Anatolian Turkey and Uromia in West Azerbaijan Province, Iran (Baran and Atatür 1998; Ananjeva et al. 2006; Sindaco and Jeremčenko 2008; Arakelyan et al. 2011; Karamiani et al. 2015).

The minor snake-eyed skink, *A. grayanus* was first described as *Blepharosteres grayanus* from Waggur District, northeast Kutch, India. Later, it was treated as a subspecies of *A. pannonicus* (Fühn 1969a). *Ablepharus grayanus* is distributed in northern and western India, through Pakistan and Afghanistan (Khan 2002) and was recorded from the eastern and south-eastern margins of the central Iranian Plateau (Fühn 1969a; Leviton and Anderson 1970), but authors, such as Anderson (1999), Rastegar-Pouyani et al. (2008) and Šmíd et al. (2014), have not listed *A. grayanus* for the Iranian herpetofauna. Karamiani et al. (2015) reported a range expansion in the Minor Snake-eyed Skink, *A. grayanus*, from eastern and south-eastern Iran, that, based on examination of pholido-

sis and morphometric measurements, were different from *A. pannonicus*.

The Asian Snake-eyed Skink, *A. pannonicus*, is distributed in a vast area from west to south in the Zagros, central Iranian Plateau and Kopet-Dagh ranges (Anderson 1999; Šmíd et al. 2014; Karamiani et al. 2018a). This species, in its extensive distribution range, based on the morphological characters, has previously been divided into several species, but later, all of them have been treated as synonyms of *A. pannonicus* (Anderson 1999; Bauer et al. 2003). So far, molecular studies for phylogenetic relationships have not been performed in species of the genus *Ablepharus* occurring in the Iranian Plateau.

In this study, we evaluate the phylogenetic relationships amongst species, based on sequences of three mitochondrial genes: COI, 12S rRNA and 16S rRNA. In addition, divergence times within the genus *Ablepharus* are estimated and a hypothesis of the historical biogeography on the Iranian Plateau is presented.

Materials and methods

Sampling and DNA extraction

The tissue samples of *Ablepharus* specimens were collected from various populations throughout the Iranian Plateau from April to October 2010 and 2019 (about 150 days in total). Moreover, we used voucher specimens in the Zoological Museum of Tehran University (ZUTC) and Zoological collection of Hakim Sabzevari University (ZCHSU). Collected specimens were preserved in 80% ethanol and were deposited in the Razi University Zoological Museum (RUZM, Kermanshah, Iran). The outgroup taxa, *Plestiodon elegans* (Boulenger, 1887) and *Ophiomorus persicus* (Steindachner, 1867) were chosen according to Pyron et al. (2013). For DNA extraction, tail tips were used, following Kapli et al. (2013). The quality of extracted DNA was measured using 1% agarose gels stained by 0.5 µl GreenViewer 6X and visualised under ultraviolet light.

The tissues and DNA samples were deposited in the Department of Biology, Hakim Sabzevari University, Sabzevar, Iran. In total, 53 samples were used for this study, including 51 ingroup and two outgroup samples (see Suppl. material 1: Table S1). We used 128 sequences of 16S rRNA for *Ablepharus* species for estimated phylogenetic relationships, including Iranian (n = 61) and Iraqi (n = 3) samples (Fig. 1) with Cyprus (n = 11), India (n = 1), Greece (n = 35), Turkey (n = 9) and Syria (n = 8) sequences retrieved from GenBank.

PCR amplification and sequencing

Three mitochondrial genes were selected for molecular phylogenetic analysis: 1) a partial sequence (631 bp) of the protein-encoding COI gene, 2) a partial sequence

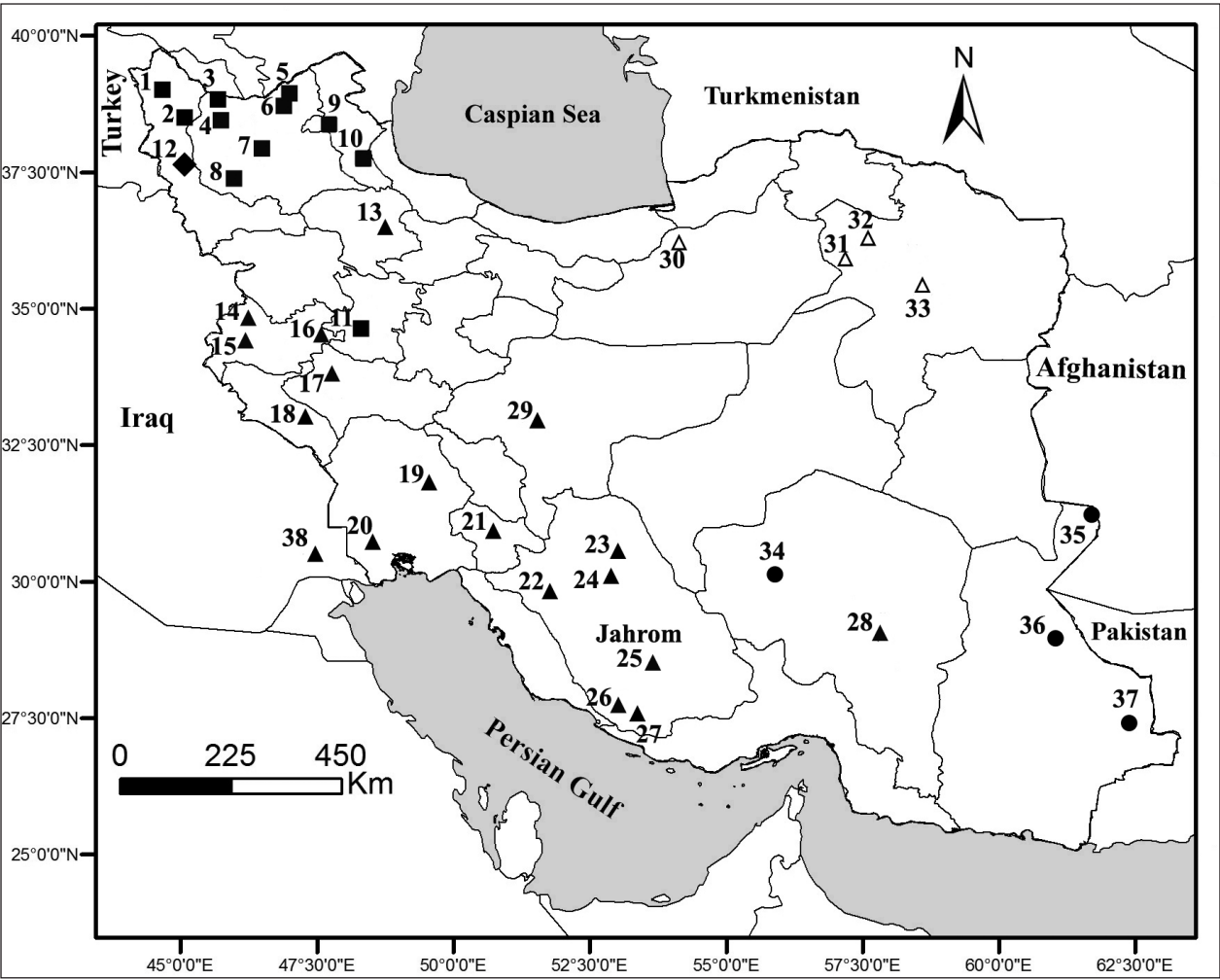


Figure 1. Map of Iran and Iraq showing the sampling localities of *Ablepharus bivittatus* (squares), *A. chernovi* (diamond), *A. grayanus* (circles), *Ablepharus* sp. (black triangles) and *A. pannonicus* (white triangles) used in this study. Numbers correspond to specimens listed in Suppl. material 1: Table S1.

Table 1. Primers used in this study to amplify and sequence marker genes.

Gene	Primer	Orientation	Primer sequence (5'→3')	Reference(s)
12S rRNA	L12S	F	AAACTGGGATTAGATACCCCACTAT	Kocher et al. 1989
	H12S	R	GAGGGTGACGGGCGGTGTGT	Kocher et al. 1989
16S rRNA	L16S	F	CGCCTGTTTATCAAAAACAT	Kocher et al. 1989
	H16S	R	CCGGTCTGAATCAGATCAGC	Kocher et al. 1989
COI	LCOI	F	TNTTMTCAACNAACCACAAAGA	Nagy et al. 2012
	HCOI	R	GGTCAACAAATCATAAAGATATTGG	Nagy et al. 2012
	H7086	R	CCTGAGAATARKGGGAATCAGTG	Nagy et al. 2012

(360 bp) of the non-coding 12S rRNA and 3) a partial sequence (532 bp) of the non-coding 16S rRNA. The genes were amplified by the polymerase chain reaction (PCR) procedure using primers presented in Table 1. PCR amplification for each of the three primers was performed in the condition of 1.5 mM MgCl₂ and 36 cycles with different temperatures and times, also for COI gene using semi-nested runs (for the semi-nested PCR run, one of the primers used in the first run was used again and the other primer was within the target sequence) (Table 2). The amplified products of three mtDNA genes were sequenced by an automated sequencer ABI 3730XL (Macrogen, Seoul, South Korea).

Alignment and genetic divergence

Alignment of the individual and concatenated COI, 12S rRNA and 16S rRNA sequences was performed with Clustal W (Thompson et al. 1994) as implemented in Bioedit version 7.0.0 (Hall 1999), then checked and corrected by eye. Sequences of COI gene were translated by the programme MEGA 7 (Tamura et al. 2013) into amino acids for checking unexpected stop codons. Uncorrected genetic distance (*p*-distance) and corrected genetic distance (K2p; Kimura 1980) were calculated using MEGA 7 (Tamura et al. 2013) for each gene in specimens from the Iranian Plateau (Tables 3–5); the concatenated COI,

Table 2. PCR programme used in this study: Primers L12S and H12S (360 bp from the 12S rRNA region), L16S and H16S (532 bp from the 16S rRNA), LCOI and HCOI (631 bp from the COI) and semi-nested (SE) for primers LCOI and H7086 (1111 bp from the COI) of the mtDNA.

PCR Step	Temperature (12S rRNA; 16S rRNA; COI; SE) [°C]	Duration (12S rRNA; 16S rRNA; COI; SE) [s]
Denature template	94 / 94 / 94 / 94	30 / 40 / 40 / 40
Anneal primers	48 / 48 / 49 / 51	45 / 40 / 40 / 40
Extension	72 / 72 / 72 / 72	60 / 80 / 90 / 90

Table 3. Pairwise divergence amongst clades derived from the 12S rRNA mitochondrial gene; p-distances (**bold**) above and K2P below diagonal. Note that the outgroup taxa are *Plestiodon elegans* and *Ophiomorus persicus*.

Clades	1	2	3	4	5	6
1. Clade A, <i>A. bivittatus</i>		0.098	0.118	0.129	0.126	0.169
2. Clade B, <i>A. chernovi</i>	0.107		0.106	0.141	0.126	0.170
3. Subclade C1a, <i>A. pannonicus</i>	0.131	0.116		0.094	0.070	0.166
4. Subclade C2: <i>Ablepharus</i> sp.	0.144	0.159	0.102		0.087	0.162
5. Subclade C1b, <i>A. grayanus</i> complex	0.141	0.139	0.075	0.094		0.156
6. Outgroup	0.195	0.197	0.190	0.184	0.177	

Table 4. Pairwise divergence amongst clades derived from the 16S rRNA mitochondrial gene; p-distances (**bold**) above and K2P below diagonal. Note that the outgroup taxa are *Plestiodon elegans* and *Ophiomorus persicus*.

Clades	1	2	3	4	5	6
1. Clade A, <i>A. bivittatus</i>		0.073	0.055	0.056	0.061	0.141
2. Clade B, <i>A. chernovi</i>	0.077		0.092	0.082	0.093	0.152
3. Subclade C1a, <i>A. pannonicus</i>	0.057	0.098		0.059	0.046	0.158
4. Subclade C2: <i>Ablepharus</i> sp.	0.059	0.087	0.062		0.060	0.157
5. Subclade C1b, <i>A. grayanus</i> complex	0.064	0.100	0.047	0.063		0.155
6. Outgroup	0.159	0.172	0.181	0.180	0.176	

Table 5. Pairwise divergence amongst clades derived from the COI mitochondrial gene; p-distances (**bold**) above and K2P below diagonal. Note that the outgroup taxa are *Plestiodon elegans* and *Ophiomorus persicus*.

Clades	1	2	3	4	5	6
1. Clade A, <i>A. bivittatus</i>		0.227	0.224	0.21	0.246	0.208
2. Clade B, <i>A. chernovi</i>	0.232		0.166	0.169	0.197	0.183
3. Subclade C1a, <i>A. pannonicus</i>	0.233	0.249		0.159	0.214	0.184
4. Subclade C2: <i>Ablepharus</i> sp.	0.217	0.256	0.185		0.197	0.197
5. Subclade C1b, <i>A. grayanus</i> complex	0.216	0.231	0.196	0.193		0.197
6. Outgroup	0.246	0.300	0.250	0.270	0.276	

12S rRNA and 16S rRNA mitochondrial gene for only the Iranian samples and 16S rRNA gene alone for combination of all samples (including the Iranian, Cyprus, India, Greece, Turkey and Syria).

We applied jModelTest 2.1.1 (Posada 2008) for extracting the best-fitting evolutionary model with Akaike Information Criterion (AIC). The best fit model for the combined dataset (COI, 12S rRNA and 16S rRNA) was TVM+I+G and that for 16S rRNA gene was TIM+I+G.

Maximum Likelihood (ML) phylogenetic tree was recovered by raxmlGUI 2.0 (Edler et al. 2020) and significance was estimated by 1000 bootstrap repartitions. Bayesian Inference (BI) was performed with the programme MrBayes 3.2.5 (Huelsenbeck and Ronquist 2001) under default for the number of runs and the num-

ber of chains for 10^7 generations and sampled every 1000 generations with 25% of the initial samples discarded as burn-in (Condamine et al. 2015). We used all snake-eyed lizard sequences for Reconstruct Ancestral State in Phylogenies (RASP) (Yu et al. 2012). The final trees were combined as a 50% majority-rule consensus tree to calculate posterior probabilities (PPs) for each node.

Estimation of divergence time

BEAST 1.8 (Heled and Drummond 2010) was used to estimate divergence times amongst the *Ablepharus* populations of this study using sequences of 16S rRNA only, because the rate for the other two genes was not available. The models and priors were applied as follows: random starting tree; clock models were set as log-normal relaxed clock with unlinked status; tree priors were set as coalescent and constant size. Finally, divergence times were assessed by the mean rate of molecular evolution for the mean rate under the uncorrelated log-normal relaxed molecular clock (ucln) priors for 16S rRNA (mean: 0.00457, standard deviation: 0.0025) (Poulakakis et al. 2005; Barley et al. 2015).

Results

Phylogenetic relationships amongst populations of *Ablepharus* in Iran and Iraq

Of the 360 bp examined for 12S rRNA gene, 67 (18.6%) were variable and 25 (6.9%) were parsimony informative. From the 532 bp of 16S rRNA, 67 (12.5%) were variable and 43 (8.1%) were parsimony informative, of 631 bp of COI 217 (34.4%) were variable and 59 (9.4%) were parsimony informative. In total 1,523 nucleotides were unambiguously aligned in a combined dataset, including 416 (28.3%) variable and 147 (9.6%) parsimony informative sites. The nucleotide frequencies in the combined dataset are as follows: A = 30%, C = 28%, G = 17% and T = 25%. P-distances and corrected genetic distances (K2P; Kimura 1980) for the individual clades are listed in Table 6. Reconstructed phylogenetic trees using COI, 12S rRNA and 16S rRNA, as well as the

Table 6. Pairwise divergence amongst clades derived from the concatenated COI, 12S rRNA and 16S rRNA mitochondrial genes; p-distances (**bold**) above and K2P below diagonal. Note that the outgroup taxa are *Plestiodon elegans* and *Ophiomorus persicus*.

Clades	1	2	3	4	5	6
1. Clade A, <i>A. bivittatus</i>		0.141	0.142	0.137	0.136	0.181
2. Clade B, <i>A. chernovi</i>	0.158		0.154	0.153	0.160	0.204
3. Subclade C1a, <i>A. pannonicus</i>	0.160	0.175		0.123	0.108	0.186
4. Subclade C2: <i>Ablepharus</i> sp.	0.154	0.174	0.136		0.120	0.194
5. Subclade C1b, <i>A. grayanus</i> complex	0.153	0.182	0.118	0.133		0.189
6. Outgroup	0.210	0.240	0.217	0.228	0.220	

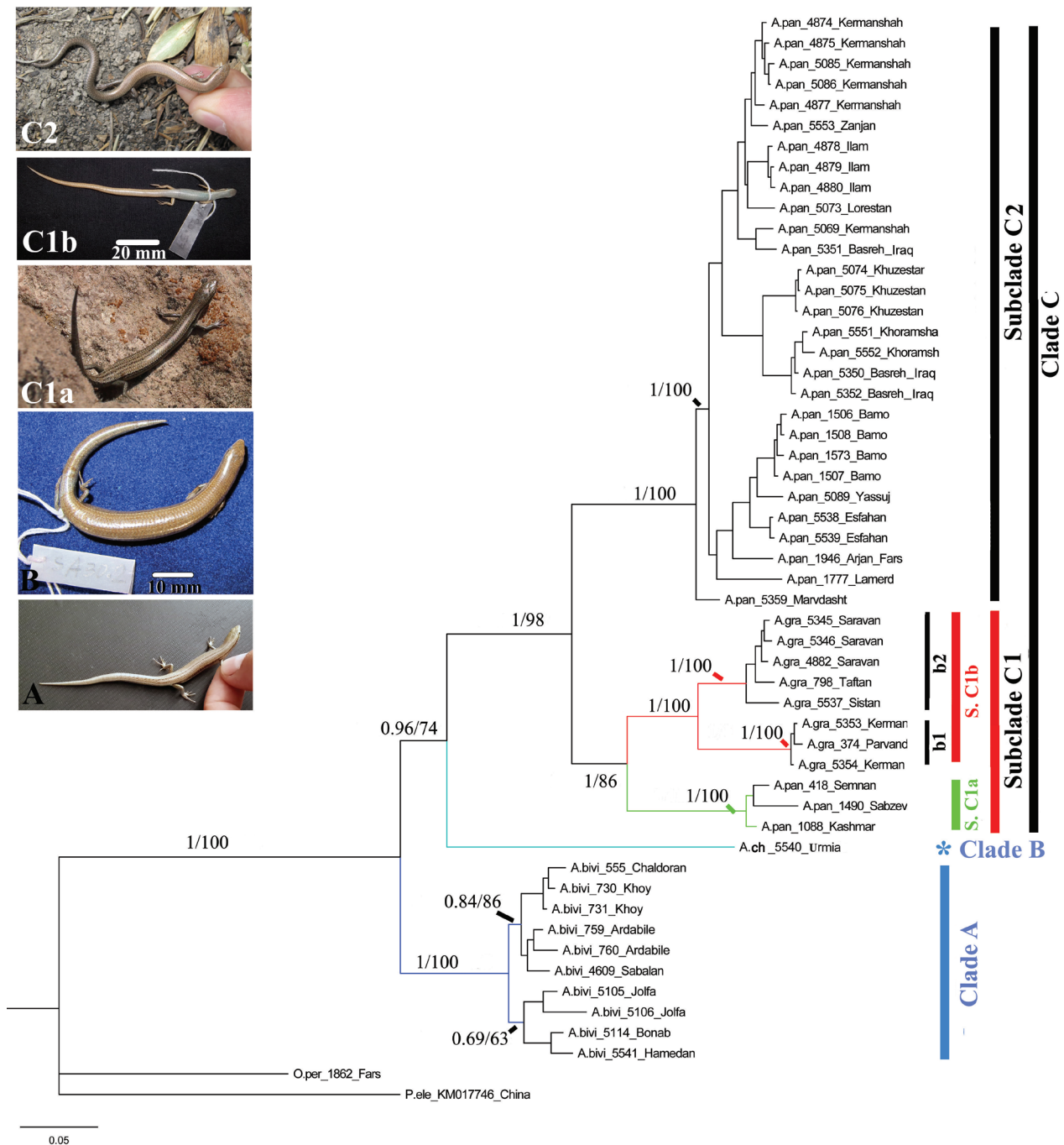


Figure 2. Cladogram of the strict-consensus tree obtained from Bayesian analysis using *Plestiodon elegans* and *Ophiomorus persicus* as outgroups. Based on the combined dataset of 1,523 bp of COI, 12S rRNA and 16S rRNA mtDNA genes. *Ablepharus bivittatus* (Clade 1); *A. chernovi* (Clade 2); *A. pannonicus* includes populations from Khorasan Razavi and Semnan Provinces (subclade C1a), *A. grayanus* complex includes populations from east and south-eastern Iran (subclade C1b) and *Ablepharus* sp. is contributed in eastern parts of Iraq and the west and southwest of Iran (subclade C2). Numbers next to the nodes are posterior probabilities followed by Maximum Likelihood (ML) bootstrap support values. Significant values are usually those over 90% for BI and over 70% for ML.

combined dataset, showed a similar topology, except in some terminal nodes. Hence, only the Bayesian tree of the combined dataset is presented in Fig. 2. Three major clades (A, B and C), recovered in all analyses, are statistically very well supported. Clade A, as a well-supported monophyletic clade, contains populations of *A. bivittatus* from Azerbaijan and Hamedan Provinces. Clade B includes *A. chernovi* from Uromia in West Azerbaijan. Its sister relationship to clade C is strongly supported by all

analyses (Fig. 2). The largest clade in the tree, clade C, is divided into subclades (C1 and C2), with subclade C1 being subdivided into lineages (C1a, C1b; Fig. 2). Clade C is very heterogeneous, contains many populations and is distributed in a vast area, from south-eastern Iraq to southwest and west of Iran, deep along the Zagros and Alborz mountains to northeast and south of the country. Lineage C1a, attributed to *A. pannonicus*, includes populations from the foothill regions of Khorasan Razavi

and Semnan Provinces, while the lineage C1b, attributed to *A. grayanus*, includes populations from eastern and south-eastern Iran. Lineage C1b was also divided into two well-supported sub-lineages (b1 and b2) of haplotypes that correspond to two geographical areas; the first (b1) consisted of the protected Parvand National Park (Sabzevar) and Kerman and the second (b2) of Sistan and Baluchistan Province. Finally, subclade C2 is distributed in the eastern part of Iraq and the west and southwest of Iran (Fig. 2). The phylogenetic relationship, produced from the molecular dataset, supports the putative new species predicted from the morphological classification of our populations, with the exception of *A. pannonicus* in Iraq and the west and southwest of Iran.

Of 536 bp examined for 16S rRNA gene, 143 (26.7%) were variable and 63 (11.8%) parsimony informative. The estimated nucleotide frequencies were: A = 33%, C = 23%, G = 20% and T = 24%. P-distances and K2P distances for the comparison amongst clades are listed in Table 7. Comparison of *Ablepharus* from Cyprus, India, Greece, Turkey and Syria with those from Iran and Iraq, based on BI in the 16S rRNA gene clearly showed that *A. chernovi* from Iran (clade B) is close to *A. chernovi* from Turkey and Syria with very low genetic distance (p-distance = 0.018). The resulting tree also indicated that populations of Khorasan Razavi and Semnan Provinces (C1a) are close to the *grayanus* complex. The *grayanus* complex forms two subclades with relatively low genetic distance (p-distance = 0.035): populations from Dostmohammad (border of Iran with Afghanistan), Kerman and west Khorasan which are closely related to the Indian single sample (b1) and populations of central areas of Sistan and Baluchistan (b2). The results showed that populations from south-eastern Iraq, western and south-western Iran are sister taxa to the remaining populations (Fig. 3). Genetic divergence between C2 and each of the other populations is more than 6% for 16S rRNA; therefore, we consider it as an undescribed species (*Ablepharus* sp.).

Biogeographical analysis

Estimated divergence times (Fig. 4) indicated that the ancestors of *Ablepharus* species diverged around 22.5 Mya and subsequently resulted in the evolution of two groups. The first group includes *A. budaki* from Cyprus, Syria

(Alawit Mountain, Aleppo, Lattakia); *A. kitaibelii* from Greece, Turkey (Izmir) and *A. chernovi* which is now distributed in a vast area in Syria (Homs), Turkey to north-western Iran (Uromia). The ancestor of *A. chernovi* diversified from *A. kitaibelii* about 13 Mya and the second group subdivided into four lineages. Divergence time of the lineages A (*A. bivittatus*), B and C is about 15.2 Mya (Fig. 4). With regard to genetic distance analysis, the most important divergence times are within the C populations. The split of the C2 (*Ablepharus* sp.) populations from the C1 populations occurred at about 10.59 Mya; about 8.41 Mya C1a (*A. pannonicus*) from C1b (*A. grayanus* complex); about 6.79 Mya C1b1 (Dostmohammad and India) from C1b2 (Kerman, Parvand Park and Sistan and Baluchistan) (Fig. 4).

Discussion

Molecular phylogeny of the genus *Ablepharus*

The results of this study show a well-resolved molecular phylogeny and identify species, based on sequence divergence of the genus *Ablepharus* in Iran and Iraq and their relationship with species existing in Cyprus, India, Greece, Turkey and Syria. Different lineages within the genus *Ablepharus* have diverged in the early Miocene and have undergone different evolutionary histories. The phylogenetic analyses (ML and BI) reveal at least five lineages of *Ablepharus* within Iran and Iraq with high bootstrap values and posterior probabilities.

A molecular phylogenetic study confirmed *A. chernovi* to represent a genetically-distinct species (Poulakakis et al. 2005). It is distributed in Armenia, Syria and Turkey (Baran and Atatür 1998; Ananjeva et al. 2006; Arakelyan et al. 2011; Sindaco et al. 2013). This species was recorded for the first time in Iran from 40 km north of Uromia, Province of West Azerbaijan, Iran by Karamiani et al. (2018b). Our analysis of three mitochondrial genes confirmed the presence of *A. chernovi* (clade B) in Iran as a sister taxon to all samples in clade C. This study confirmed previous records of *A. grayanus* in Iran and indicated a high level of variation in populations of eastern and south-eastern Iran. In addition, analysis of the 16S rRNA gene revealed a close

Table 7. Comparative pairwise divergence amongst all clades in Iran, Iraq, Cyprus, India, Greece, Turkey and Syria derived from the 16S rRNA mitochondrial gene; p-distances (**bold**) above and K2P below diagonal.

Clades	Clade_1	Clade_2	Clade_3	Clade_4	Clade_5	Clade_6	Clade_7	Clade_8	Clade_9
Clade_1		0.079	0.075	0.065	0.076	0.163	0.068	0.072	0.066
Clade_2	0.084		0.111	0.082	0.108	0.155	0.086	0.052	0.074
Clade_3	0.079	0.120		0.068	0.058	0.188	0.104	0.107	0.110
Clade_4	0.068	0.087	0.071		0.060	0.183	0.095	0.085	0.097
Clade_5	0.081	0.117	0.061	0.063		0.186	0.103	0.106	0.156
Clade_6	0.186	0.175	0.219	0.213	0.218		0.159	0.175	0.156
Clade_7	0.072	0.091	0.113	0.103	0.112	0.180		0.064	0.055
Clade_8	0.075	0.054	0.116	0.091	0.115	0.201	0.068		0.063
Clade_9	0.069	0.078	0.119	0.105	0.119	0.176	0.058	0.067	



herpetozoa.pensoft.net

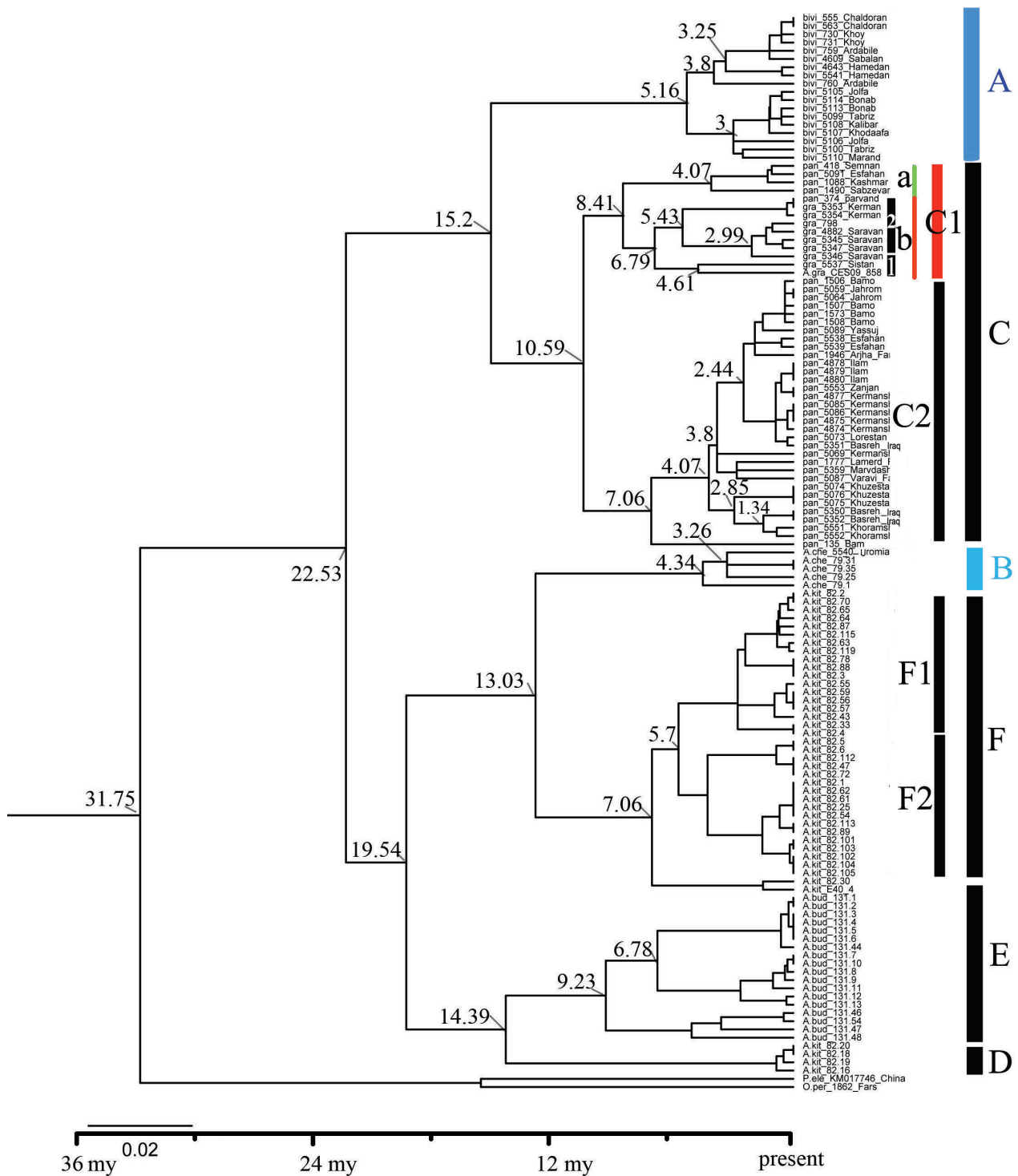


Figure 4. Chronogram of BEAST analysis using only 16S rRNA for specimens of Iran, Cyprus, India, Greece, Turkey and Syria.

relationship between populations of east Iran (5537 Dostmohamad) and India (CES09-858). Our molecular results showed that *A. pannonicus* is a paraphyletic taxon regarding the position of *A. grayanus* in the phylogenetic tree (Fig. 2). Therefore, to solve the taxonomic problems of *A. pannonicus* in its distribution area, the population of subclade C1a from northeast should be considered as *A. pannonicus*, because of its relatively

short geographic distance to the type population. The populations of subclade C2 should be considered as a separate evolutionary species. *Ablepharus bivittatus* (clade A) and *A. chernovi* (clade B) are monophyletic units, supporting the taxonomic status of these species. Although there was only one sample available for *A. chernovi*, it is quite divergent from the other clades in all analyses (Table 6).

Biogeography of the genus *Ablepharus*

Fühn (1969a) believed that the representative distribution is a good reason to accept the centre of origin of *Ablepharus* in SE Asia, but the fossil fragment of a jaw found in lower Pleistocene layer, extends the ancient range of *Ablepharus* to 800 km northwards from the Altai in Kazakhstan (Fühn 1969b; Anderson 1999). The past and current model for distribution and habitat suitability for *Ablepharus* confirmed that they prefer different climatic conditions across the Middle East, Central Asia (Tajikistan, Turkmenistan and Uzbekistan) to India (Karamiani et al. 2018a). Based on the fossil evidence from Altai (south of Russia, close to the border with Kazakhstan; Estes 1983), we suggest that Central Asia is the origin of the genus *Ablepharus*. After the rise of Kopet Dag, Caucasus and Zagros Mountains, they have subsequently been dispersed into the Iranian Plateau. The route of its dispersal is suggested to be similar with *Paralaudakia caucasia* (Macey et al. 1998) and in the opposite direction as *Eremias velox*, which dispersed from the Iranian Plateau to Central Asia before the rise of the Caucasus Mountains (Rastegar-Pouyani 2009; Rastegar-Pouyani et al. 2012). According to Brandley et al. (2011, 2012), *Plestiodon*, the outgroup taxon of this study, originated from Eastern Eurasia. *Plestiodon* was separated from the ancestor of *Ablepharus* soon after collision of the Indian Plate to Eurasia and the rise of the Himalaya Mountains and the Tibetan Plateau around Oligocene (32 Mya) (Tripathy-Lang et al. 2013). In addition, the collision of the Indian Plate with Eurasia resulted in changing the geological structure of Iran, so that the ocean between the Lut Block and the Afghan Block was closed (Aghanabati 2004). During the Eocene to the early Miocene (55 to 20 Mya), at the same time as the formation of the Red Sea, with the collision of Africa with Europe, a new Alpine orogeny was introduced into Europe and the new Tethys Sea disappeared (Darvishzadeh 2003; Berra and Angiolini 2014; Karamiani 2016). Based on our molecular clock, two groups of the snake-eye lizard had diverged about 22.5 Mya and subsequently dispersed into different areas. The first group occurs in south-eastern Europe, Greece, Turkey, north-western Iran, Cyprus and Syria and the second group on the Iranian Plateau and its adjoining eastern regions. Recently, fossil remains of skinks have been described (e.g. *Ablepharus*) from Greece (late Miocene and early Pliocene) and from the European part of Turkey (Georgalis et al. 2019, 2021; Loréal et al. 2020; Villa et al. 2020). For the distribution of taxa of the first group, two routes are assumed: one is dispersal to south-eastern Europe and then to southern Europe and the latter to the Alps, Greece, Turkey and Syria about 19.5 Mya. The second route is proposed to the north of Iran through Kopet Dag Mountains then to the Caucasus Mountains and finally to Southeast Europe, Greece, Turkey, Cyprus and Syria. The second route (northern of Iran) seems more likely because the ancient Tethys Sea has been an ob-

stacle to the dispersal of taxa to European regions. The Asia-Anatolian route (the second route) is also suggested by Poulakakis et al. (2005).

Our results showed that, amongst the Iranian Plateau taxa, clade B lies within the populations of south-eastern Europe, Greece, Turkey, Cyprus and Syria and is a sister taxon to *A. chernovi* from Turkey. This clade divided from *A. kitaibelii* about 13 Mya. Most likely, *A. chernovi* reached Iran following its dispersal through the Zagros Mountain belt in south-eastern Turkey and it seems that Uromia Lake was a barrier against its dispersal. Perhaps in higher regions, such as Khoi and Chaldoran, it could not expand because its favourable niches in the area are occupied by *A. bivittatus* (probably having similar ecological niches). Around the middle Miocene period (about 15 Mya), much of the Tethys Sea in Iran retreated (Darvishzadeh 2003). The Iranian populations have mainly been divided into two groups: the first group, the clades A and C in this study. Collision of the Arabian Plate to the Iranian Plateau resulted in raising the Zagros, Caucasus and Kopet Dag mountains. The current distribution of clade A (attributed to the species of *A. bivittatus*) is confined to the Caucasus Mountains and west of the Kopet Dag and south of Turkmenistan (Anderson 1999; Karamiani et al. 2017). In the late Miocene, about 10 Mya, the rise of the Zagros Mountains (Sborshchikov et al. 1981) caused the splitting of clade C into two branches, subclade 1 and subclade 2. About 8 Mya, Helmand Zone and the eastern and northeast Iranian Plateau were the habitat of ancestors of subclade 1 and about 7 Mya, progenitors of subclade 2 occurred in the west and southwest Zagros Mountains. The subclade 1 has been dispersed in east and southeast Iran, Afghanistan, Pakistan, north and west India and Central Asia (the Pamir-Altaï Mountains). Subclade 2 dispersed into the west, south and southwest Zagros Mountains (Anderson 1999; Khan 2002; Karamiani 2016). The Arabian Plate separated from Africa at the beginning of Pliocene (about 5 Mya; Girdler 1984) and this phenomenon accelerated uplifting of Caucasus and Kopet Dag mountains in the north and the Zagros Mountains in the south of the Iranian Plateau. Almost coincident with the rise of the Zagros, the populations of Khuzestan, Iran and Basra, Iraq dispersed into other regions of west and southwest Iran. The distribution area of the lineage C1a (population of north-eastern Iran) reaches the Shahrood Fault, which is the northern boundary of the eastern and south-eastern part of the Iranian Plateau (Darvishzadeh 2003). The ancestor of sub-lineage C1b separated into two groups around 5.4 Mya in Kerman-Parvand and Saravan-Taftan (Sistan and Baluchistan) areas. The drying Neotethys Sea in the Pliocene (Aghanabati 2004) introduced divergence in populations of south-eastern Iran. The global climate in the late Miocene has changed from moist to dry (Janis 1993), causing the drying of Asia (Guo et al. 2004). Climate change and changing habitats may have a fundamental role in speciation and widespread extinction (Hewitt 1996; He et al. 2010). Considering the divergence

time of clade A (*A. bivittatus*) from its central Asian ancestor (about 5 Mya), it may have occurred after orogeny of the Caucasus, Kopet-Dagh and Alborz mountains. It has subsequently dispersed to the northwest of Iran and then dispersed to the Zagros Mountains. The warm and humid climate (3.6–2.5 Mya; Webb and Bartlein 1992) led to the fragmentation of plant and animal populations (Bennett 1990). It can be assumed that, at this time, most populations of *Ablepharus* had invaded or sheltered in cool habitats (e.g. migration of *A. bivittatus* from northwest to lower latitudes, such as Hamadan; 3.25 Mya). The climate changed to cool and dry (Webb and Bartlein 1992) in the early Pleistocene (2.5–1.8 Mya). The snow line in the cold weather periods in Hamadan has been up to 1800 m (Wright 1962), so it is likely that the Hamadan population was sheltered at low altitudes. The effect of hot and humid climate (6.5–2.5 Mya) on subclade 2 caused divergence of Zagros (Baghmalek, north Khuzestan) from Abadan (south Khuzestan), Iran and Basra clades in Iraq about 2.8 Mya. The Northern Zagros clade (Zanjan, Kermanshah, Ilam and Lorestan) has diverged from central and southern Zagros populations (Kohkiluyeh and Boyer Ahmad, Isfahan and Fars areas) about 2.4 Mya.

Acknowledgements

The authors thank Prof. Steven C. Anderson (University of the Pacific, Stockton, California, USA) and Prof. Theodore J. Papenfuss (University of California, USA) for their kind help to improve the English of our manuscript. We thank Prof. Alireza Sari and Hassan Salehi from Zoological Museum, University of Tehran, for the loan of some specimens of *Ablepharus*. We thank Seyyed Saeed Hosseini Yousefkhani for his help in molecular laboratory and photo of C1a (Fig. 2), Akbar Fattahi, Mousa Mahmoodi, Ali Gholamifard, Hiwa Faizi, Mohsen Takesh, Ehsan Damadi, Morteza Akbarpour and Manocher Mohammadi for assisting us with fieldwork in Iran. We thank Fadhil Abbas Rhadi and Rihab Ghaleb Mohammed for supplying specimens in Iraq. The molecular analyses were performed in High Performance Computing Research Centre (HPCRC) of Amirkabir University.

References

- Aghanabati SA (2004) Geology of Iran. Geological Survey of Iran Publication, Tehran, 586 pp.
- Ananjeva NB, Orlov NL, Roman GK, Darevsky SD, Ryabov SA, Barabanov AV (2006) The reptiles of northern Eurasia – taxonomic diversity, distribution, conservation status. Zoological Institute, Russian Academy of Sciences, Pensoft Series Faunistica, No. 47, Sofia, 246 pp.
- Anderson SC (1999) The Lizards of Iran. Society for the Study of Amphibians and Reptiles, Oxford, Ohio, 442 pp.
- Arakelyan MS, Danielyan FD, Corti C, Sindaco R, Leviton AE (2011) Herpetofauna of Armenia and Nagorno-Karabakh. Society for the Study of Amphibians and Reptiles, Oxford, Ohio, 149 pp.
- Baran İ (1977) Türkiye’de Scincidae familyası türlerinin taksonomisi. Doğa Bilim Dergisi 1: 217–223.
- Baran I, Atatür MK (1998) Turkish herpetofauna (amphibians and reptiles). Republic of Turkey Ministry of Environment, Ankara, 214 pp.
- Barley AJ, Datta-Roy A, Karanth KP, Brown RM (2015) Sun skink diversification across the Indian–Southeast Asian biogeographical interface. Journal of Biogeography 42: 292–304. <https://doi.org/10.1111/jbi.12397>
- Bauer AM, Shea G, Günther R (2003) An annotated catalogue of the types of scincid lizards (Reptilia, Squamata, Scincidae) in the collection of the Museums für Naturkunde der Humboldt-Universität zu Berlin (ZMB). Zoosystematics and Evolution 79: 253–321. <https://doi.org/10.1002/mmzn.20030790204>
- Bennett K (1990) Milankovitch cycles and their effects on species in ecological and evolutionary time. Paleobiology 16: 11–21. <https://doi.org/10.1017/S0094837300009684>
- Berra F, Angiolini L (2014) The Evolution of the Tethys Region throughout the Phanerozoic: A Brief Tectonic Reconstruction. In: Marlow L, Kendall CCG, Yose LA (Eds) Petroleum systems of the Tethyan region, American Association of Petroleum Geologists, 27 pp.
- Bibron M, Bory de Saint-Vincent JP (1833) Vertébrés a sang froid. Reptiles et poissons. Reptiles. In: Geoffroy & Geoffroy, Expédition Scientifique de Morée, Zoologie 3: 69–70.
- Blanford WT (1876) The Zoology and Geology. In: Blanford WT (Ed.) Eastern Persia, an Account of the Journeys of the Persian Boundary Commission, 1870–1872 (Vol. 2). Macmillan and Co., London, 516 pp.
- Boulenger GE (1887) Catalogue of the lizards in the British Museum (Natural History), 3, Lacertidae, Gerrhosauridae, Scincidae, Anelytropidae, Dibamidae, Chamaeleontidae. British Museum (Natural History), London, 575 pp.
- Brandley MC, Wang Y, Guo X, de Oca ANM, Fería-Ortiz M, Hikida T, Ota H (2011) Accommodating heterogeneous rates of evolution in molecular divergence dating methods: an example using intercontinental dispersal of *Plestiodon* (*Eumeces*) lizards. Systematic Biology 60: 3–15. <https://doi.org/10.1093/sysbio/syq045>
- Brandley MC, Ota H, Hikida T, Nieto Montes De Oca A, Fería-Ortiz M, Guo X, Wang Y (2012) The phylogenetic systematics of blue-tailed skinks (*Plestiodon*) and the family Scincidae. Zoological Journal of the Linnean Society 165: 163–189. <https://doi.org/10.1111/j.1096-3642.2011.00801.x>
- Condamine FL, Nagalingum NS, Marshall CR, Morlon H (2015) Origin and diversification of living cycads: a cautionary tale on the impact of the branching process prior in Bayesian molecular dating. BMC Evolutionary Biology 15: 1–65. <https://doi.org/10.1186/s12862-015-0347-8>
- Darevsky IS (1953) *Ablepharus chernovi* spec. nov. (Reptilia, Sauria) in Armenian Republic [in Russian]. Bulletin de la Société des Naturalistes Moscou 58: 39–41.
- Darvishzadeh A (2003) Geology of Iran. Amirkabir Publication, Tehran, 902 pp.
- Edler D, Klein J, Antonelli A, Silvestro D (2020) raxmlGUI 2.0: a graphical interface and toolkit for phylogenetic analyses using RAxML. Methods in Ecology and Evolution 12: 1–5. <https://doi.org/10.1101/800912>
- Eremchenko VK, Szczerbak NN (1986) *Ablepharus* lizards in the fauna of the USSR and neighbouring countries. Frunze (Akad. Nauk. Kirgis. SSR), 171 pp.
- Estes R (1983) Sauria Terrestria, Amphisbaenia. In: Wellnhofer P (Ed.) Encyclopedia of Paleoherpétology, part 10a. Gustav Fischer Verlag, Stuttgart – New York, 249 pp.

- Fitzinger L (1823) in Lichtenstein: in Evermann's Reise [von Orenburg] nach Buchara. EHG Christiani, Berlin.
- Fitzinger L (1824) Über *Ablepharus pannonicus*, eine neue Eidechse aus Ungarn. Verhandlungen der Gesellschaft Naturforschender Freunde in Berlin 1: 297–302.
- Fühn IE (1969a) Revision and redefinition of the genus *Ablepharus* Lichtenstein, 1823 (Reptilia, Scincidae). Revue Roumaine de Biologie, ser. Zoology 14: 23–41.
- Fühn IE (1969b) The Polyphyletic origin of the genus *Ablepharus* (Reptilia, Scincidae): a case of parallel evolution. Journal of Zoological Systematics and Evolutionary Research 7: 67–76. <https://doi.org/10.1111/j.1439-0469.1969.tb00849.x>
- Georgalis G, Villa A, Martin I, Vasilyan D, Delfino M (2019) Fossil amphibians and reptiles from the Neogene locality of Maramena (Greece), the most diverse European herpetofauna at the Miocene/Pliocene transition boundary. Palaeontologia Electronica 22: 1–99. <https://doi.org/10.26879/908>
- Georgalis GL, Čerňanský A, Mayda S (2021) Late Paleogene herpetofaunas from the crossroads between two continents—new amphibian and reptile remains from the Oligocene of southern Balkans and Anatolia. Comptes Rendus Palevol 20: 253–272. <https://doi.org/10.5852/cr-palevol2021v20a15>
- Girdler RW (1984) The evolution of the Gulf of Aden and Red Sea in space and time. Deep Sea Research Part A. Oceanographic Research Papers 31: 747–762. [https://doi.org/10.1016/0198-0149\(84\)90039-6](https://doi.org/10.1016/0198-0149(84)90039-6)
- Göçmen B, Kumlutaş Y, Tosunoğlu M (1996) A new subspecies, *Ablepharus kitaibelii* (Bibron & Borry, 1833) *budaki* n. ssp. (Sauria: Scincidae) from the Turkish Republic of Northern Cyprus. Turkish Journal of Zoology 20: 397–406.
- Gray JE (1839) Catalogue of the slender-tongued saurians, with descriptions of many new genera and species. Annals and Magazine of Natural History 2: 331–337. <https://doi.org/10.1080/00222933909512395>
- Greer AE (1970) A subfamilial classification of scincid lizards. Bulletin of the Museum of Comparative Zoology 139: 151–183.
- Guo Z, Peng S, Hao Q, Biscaye PE, An Z, Liu T (2004) Late Miocene–Pliocene development of Asian aridification as recorded in the Red-Earth Formation in northern China. Global and Planetary Change 41: 135–145. <https://doi.org/10.1016/j.gloplacha.2004.01.002>
- Hall TA (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic acids symposium series 41: 95–98.
- He K, Li YJ, Brandley MC, Lin LK, Wang YX, Zhang YP, Jiang XL (2010) A multi-locus phylogeny of Nectogalini shrews and influences of the paleoclimate on speciation and evolution. Molecular Phylogenetics and Evolution 56: 734–746. <https://doi.org/10.1016/j.ympev.2010.03.039>
- Heled J, Drummond AJ (2010) Bayesian Inference of Species Trees from Multilocus Data. Molecular Biology and Evolution 27: 570–580. <https://doi.org/10.1093/molbev/msp274>
- Hewitt GM (1996) Some genetic consequences of ice ages, and their role in divergence and speciation. Biological Journal of the Linnean Society 58: 247–276. <https://doi.org/10.1006/bjil.1996.0035>
- Huelsenbeck JP, Ronquist F (2001) MRBAYES: Bayesian inference of phylogenetic trees. Bioinformatics 17: 754–755. <https://doi.org/10.1093/bioinformatics/17.8.754>
- Ilgaz Ç, Kumlutaş Y, Olgun K, Baran İ (2007) The morphology and distribution of *Ablepharus bivittatus* (Menetries, 1832) (Reptilia: Sauria: Scincidae) in Turkey. Russian Journal of Herpetology 14: 91–97.
- Janis CM (1993) Tertiary mammal evolution in the context of changing climates, vegetation, and tectonic events. Annual Review of Ecology and Systematics 24: 467–500. <https://doi.org/10.1146/annurev.es.24.110193.002343>
- Jeremčenko VK, Panfilov AM (1990) *Ablepharus darvazi* sp. nov. – a new species *Ablepharus* (Sauria, Scinsidae [Scincidae]) from Tadzhikistan. Izvestiya Akademii Nauk Kirgizskoi Ssr Khimiko-Tekhnologicheskije Biologicheskije Nauki, Frunze 4: 56–63.
- Kapli P, Botoni D, Ilgaz C, Kumlutaş Y, Avcı A, Rastegar-Pouyani N, Fathinia B, Lymberakis P, Ahmadzadeh F, Poulakakis N (2013) Molecular phylogeny and historical biogeography of the Anatolian lizard *Apathya* (Squamata, Lacertidae). Molecular Phylogenetics and Evolution 66: 992–1001. <https://doi.org/10.1016/j.ympev.2012.12.002>
- Karamiani R (2016) Systematics and phylogeny of the Snake-eyed skinks of the genus *Ablepharus* Fitzinger, 1823 (Sauria: Scincidae) in Iran using morphological and molecular data. PhD Thesis, Razi University, Kermanshah, Iran.
- Karamiani R, Rastegar-Pouyani N, Rastegar-Pouyani E, Akbarpour M, Damadi E (2015) Verification of the Minor Snake-eyed Skink, *Ablepharus grayanus* (Stoliczka, 1872) (Sauria: Scincidae), from Iran. Zoology in the Middle East 61: 226–230. <https://doi.org/10.1080/09397140.2015.1069241>
- Karamiani R, Rastegar-Pouyani N, Rastegar-Pouyani E, Dehghani A, Banan-Khojasteh SM, Mahmoodi M (2017) Phenotypical variation and range extension distribution of the Snake-eyed Skink, *Ablepharus bivittatus* (Sauria: Scincidae) in Iran. Iranian Journal of Animal Biosystematics 13: 289–294.
- Karamiani R, Rastegar-Pouyani N, Rastegar-Pouyani E (2018a) Modeling the past and current distribution and habitat suitability for *Ablepharus grayanus* and *A. pannonicus* (Sauria: Scincidae). Asian Herpetological Research 9: 56–64. <https://doi.org/10.5772/intechopen.82476>
- Karamiani R, Rastegar-Pouyani N, Rastegar-Pouyani E, Mamaghani Shishvani M, Nilsson N (2018b) First record of *Ablepharus chernovi* Darevsky, 1953, from Iran. Herpetozoa 31: 94–97.
- Khan MS (2002) Key and checklist to the lizards of Pakistan (Reptilia: Squamata: Sauria). Herpetozoa 15: 99–119.
- Kimura M (1980) A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. Journal of Molecular Evolution 16: 111–120. <https://doi.org/10.1007/BF01731581>
- Kocher TD, Thomas WK, Meyer A, Edwards SV, Pääbo S, Villablanca FX, Wilson AC (1989) Dynamics of mitochondrial DNA evolution in animals: amplification and sequencing with conserved primers. Proceedings of the National Academy of Sciences 86: 6196–6200. <https://doi.org/10.1073/pnas.86.16.6196>
- Leviton AE, Anderson SC (1970) The amphibians and reptiles of Afghanistan, a checklist and key to the herpetofauna. Proceedings of the California Academy of Sciences 38: 163–206.
- Loréal E, Villa A, Georgalis G, Delfino M (2020) Amphibians and reptiles from the late Miocene and early Pliocene of the Ptolemais area (Western Macedonia, Greece). Annales de Paléontologie 106: e102407. <https://doi.org/10.1016/j.annpal.2020.102407>
- Macey JR, Schulte JA, Ananjeva NB, Larson A, Rastegar-Pouyani N, Shammakov SM, Papenfuss TJ (1998) Phylogenetic relationships among agamid lizards of the *Laudakia caucasia* species group: testing hypotheses of biogeographic fragmentation and an area cladogram for the Iranian Plateau. Molecular Phylogenetics and Evolution 10: 118–131. <https://doi.org/10.1006/mpev.1997.0478>

- Ménétries E (1832) Catalogue Raisonné des Objets de Zoologie Recueillis dans un Voyage au Caucase et Jusqu'aux Frontiers Actuelles de la Perse Entrepris par ordre de S. M. L'Empereur. L'Académie Impériale des Sciences, St. Pétersbourg, 271 pp. <https://www.biodiversitylibrary.org/page/41529538>
- Nagy ZT, Sonet G, Glaw F, Vences M (2012) First large-scale DNA barcoding assessment of reptiles in the biodiversity hotspot of Madagascar, based on newly designed COI primers. *PLoS One* 7: e34506. <https://doi.org/10.1371/journal.pone.0034506>
- Posada D (2008) jModelTest: phylogenetic model averaging. *Molecular Biology and Evolution* 25: 1253–1256. <https://doi.org/10.1093/molbev/msn083>
- Poulakakis N, Lymberakis P, Tsigonopoulos CS, Magoulas A, Mylonas M (2005) Phylogenetic relationships and evolutionary history of snake-eyed skink *Ablepharus kitaibelii* (Sauria: Scincidae). *Molecular Phylogenetics and Evolution* 34: 245–256. <https://doi.org/10.1016/j.ympev.2004.10.006>
- Pyron R, Burbrink F, Wiens J (2013) A phylogeny and revised classification of Squamata, including 4161 species of lizards and snakes. *BMC Evolutionary Biology* 13: 1–54. <https://doi.org/10.1186/1471-2148-13-93>
- Rastegar-Pouyani E (2009) The phylogeny of the *Eremias velox* complex of the Iranian Plateau and Central Asia (Reptilia, Lacertidae): Molecular evidence from ISSR-PCR fingerprints. *Iranian Journal of Animal Biosystematics* 5: 33–46.
- Rastegar-Pouyani N, Kami HG, Rajabzadeh M, Shafiei S, Anderson SC (2008) Annotated checklist of amphibians and reptiles of Iran. *Iranian Journal of Animal Biosystematics* 4: 43–66.
- Rastegar-Pouyani E, Nourini SK, Rastegar-Pouyani N, Joger U, Wink M (2012) Molecular phylogeny and intraspecific differentiation of the *Eremias velox* complex of the Iranian Plateau and Central Asia (Sauria, Lacertidae). *Journal of Zoological Systematics and Evolutionary Research* 50: 220–229. <https://doi.org/10.1111/j.1439-0469.2012.00662.x>
- Sborshchikov IM, Savostin LA, Zonenshain LP (1981) Present plate tectonics between Turkey and Tibet. *Tectonophysics* 79: 45–73. [https://doi.org/10.1016/0040-1951\(81\)90232-8](https://doi.org/10.1016/0040-1951(81)90232-8)
- Schmidtler JF (1997) Die *Ablepharus kitaibelii*-Gruppe in Süd-Anatolien und benachbarten Gebieten. *Herpetozoa* 10: 35–63.
- Sindaco R, Jeremčenko VK (2008) The reptiles of the Western Palearctic. Annotated checklist and distributional atlas of the turtles, crocodiles, amphisbaenians and lizards of Europe, North Africa, Middle East and Central Asia. *Latina* (Edizioni Belvedere), 580 pp.
- Sindaco R, Venchi A, Grieco C (2013) The Reptiles of the western Palearctic. 2. Annotated checklist and distributional atlas of the snakes of Europe, North Africa, Middle East and Central Asia, with an update to the Vol. 1. *Monografie della Societas Herpetologica Italica*, Edizioni Belvedere, 543 pp.
- Skourtanioti E, Kapli P, Ilgaz Ç, Kumlutaş Y, Avcı A, Ahmadzadeh F, Crnobrnja-Isailović J, Gherghel I, Lymberakis P, Poulakakis N (2016) A reinvestigation of phylogeny and divergence times of the *Ablepharus kitaibelii* species complex (Sauria, Scincidae) based on mtDNA and nuDNA genes. *Molecular Phylogenetics and Evolution* 103: 199–214. <https://doi.org/10.1016/j.ympev.2016.07.005>
- Šmíd J, Moravec J, Kodym P, Kratochvíl L, Hosseinian Yousefkhani SS, Frynta D (2014) Annotated checklist and distribution of the lizards of Iran. *Zootaxa* 3855: 1–97. <https://doi.org/10.11646/zootaxa.3855.1.1>
- Steindachner F (1867) Herpetologische notizen. *Sitzungsberichte der Akademie der Wissenschaften in Wien* 55: 263–273.
- Stoliczka F (1872) Notes on the Reptilian and Amphibian Fauna of Kachh. *Proceedings of the Asiatic Society of Bengal* 1872: 71–85.
- Strauch A (1868) Ueber die Arten der Eidechsen-Gattung *Ablepharus* (Fitz.). *Bulletin de l'Académie impériale des sciences de St.-Petersbourg* 6: 553–570.
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S (2013) MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Molecular Biology and Evolution* 30: 2725–2729. <https://doi.org/10.1093/molbev/mst197>
- Thompson JD, Higgins DG, Gibson TJ (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic acids research* 22: 4673–4680. <https://doi.org/10.1093/nar/22.22.4673>
- Tripathy-Lang A, Hodges KV, van Soest MC, Ahmad T (2013) Evidence of pre-Oligocene emergence of the Indian passive margin and the timing of collision initiation between India and Eurasia. *Lithosphere* 5: 501–506. <https://doi.org/10.1130/L273.1>
- Uetz P, Hošek J (2020) The Reptile database. <http://www.reptile-database.org/>
- Villa A, Georgalis GL, Delfino M (2020) The latest early Pleistocene amphibians and reptiles from Kaias (Greece) and the first record of fossil *Ophiomorus* (Squamata, Scincidae). *Geobios* 62: 79–90. <https://doi.org/10.1016/j.geobios.2020.06.010>
- Vitt LJ, Caldwell JP (2013) *Herpetology: an Introductory Biology of Amphibians and Reptiles*. Academic Press, San Diego, 757 pp. <https://doi.org/10.1016/B978-0-12-386919-7.00002-2>
- Vyas R (2011) Preliminary Survey on Reptiles of Jassore Wildlife Sanctuary, Gujarat State, India. *Russian Journal of Herpetology* 18: 210–214.
- Webb T, Bartlein P (1992) Global changes during the last 3 million years: climatic controls and biotic responses. *Annual Review of Ecology and Systematics* 23: 141–173. <https://doi.org/10.1146/annurev.es.23.110192.001041>
- Wettstein O (1960) Lacertilia aus Afghanistan. *Contribution à l'étude de la faune d'Afghanistan* 3. *Zoologischer Anzeiger* 165: 58–63.
- Wright HE (1962) Pleistocene glaciations in Kurdistan. *Eiszeitalter und Gegenwart* 12: 31–164.
- Yu Y, Harris AJ, He XJ (2012) RASP (Reconstruct Ancestral State in Phylogenies) 2.1b. <http://mnh.scu.edu.cn/soft/blog/RASP>

Supplementary material 1

Table S1

Authors: Rasoul Karamiani¹, Nasrullah Rastegar-Pouyani¹, Eskandar Rastegar-Pouyani

Data type: table

Explanation note: Specimens used in this study (the square indicated CO1, 12S and 16S rRNA mitochondrial sequence genes; the circle revealed 16S rRNA gene). Star indicate numbers correspond to specimens listed in Figure 1.

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/herpetozoa.34.e66338.suppl1>